

Neurobehavioural and Neurochemical Changes in Arsenic Induced Cerebellar Toxicity in Male Sprague-Dawley Rats: An Experimental Study

RAVI SHANKAR PRASAD SAWAN¹, SRIDEVI NANGALI SRINIVASA², SHASHIDHAR KURPAD NAGARAJ³



ABSTRACT

Introduction: Sodium arsenite, an inorganic arsenic, is naturally present at high level (>50 µg/L) in ground water. Drinking ground water is the biggest threat to public health. Though, there are numerous reports on arsenic neurotoxicity, the arsenic effect on cerebellar neurotoxicity remains vague especially its chronic effect on its neurobehavioural and neurochemical alterations.

Aim: To evaluate the neurobehavioural and neurochemical alterations caused by sodium arsenite in cerebellum of rats.

Materials and Methods: This experimental study was conducted in the Central Animal House at Sri Devaraj Urs Academy of Higher Education and Research (SDUAHER) from November 2019 to February 2020 for a period of 90 days. Total 16 male sprague-dawley rats were randomised into two equal groups. Group I: Control, received normal saline. Group II: Sodium arsenite, doses of 50 Parts per Millions (PPM) for 90 days through oral gavage. Rats were subjected to Open Field Test (OFT) for locomotor and exploratory behaviour and Beam Walking Test (BWT) for motor coordination and balance. Following behavioural tests, rats were anaesthetised. Blood was drawn from a retro-orbital puncture. Brains were dissected and cerebellum was separated. Concentration of Malondialdehyde (MDA), Nitric Oxide (NO) and activity of Glutathione Peroxidase (GPx) were assessed spectrophotometrically in serum and cerebellum of rats. Mean±SD was used for normally distributed data and groups were compared using independent t-test, whereas for non-normally

distributed data, Median (25th-75th Percentile) was used and Mann-Whitney U-test was used to compare groups.

Results: Arsenic-treated rats showed a significant increase in arsenic concentration in serum and cerebellum (5.5±1.6 ng/mL, 2.76±0.56 µg/g, respectively) compared to control (1.14±0.43 ng/mL, 0.65±0.29 µg/g, respectively). There was a significant decrease in locomotor and exploratory behaviour and impairment in motor coordination and balance in arsenic treated rats with a p-value <0.001 in comparison with control rats. The arsenic treated rats had significantly enhanced concentration of MDA and NO level and reduced activity of GPx in serum {16.84 (13.84-18.87), 33.79 (30.05-37.17) nmol/mL, and 6.89 (5.24-8.5) mmoles of Reduced glutathione (GSH) oxidised/min/mL, respectively} compared to control {8.81 (8.36-9.48), 17.66 (15.33-21.29) nmol/mL, and 15.16 (12.77-16.59) mmoles of GSH oxidised/min/mL, respectively} and also found increased concentration of MDA and NO level and reduced activity of GPx in tissue {7.98 (7.14-8.92), 24.67 (21.4-28-22) nmol/mg of protein and 2.66 (1.19-3.86) mmoles of GSH oxidised/min/mg protein, respectively} compared to control {3.02 (2.35-3.61), 13.93 (11.0-16.16) nmol/mg of protein and 7.63 (7.08-9.19) mmoles of GSH oxidised/min/mg protein, respectively}.

Conclusion: The oral administration of sodium arsenite at the doses of 50 PPM for 90 days showed interesting alterations in neurobehavioural and neurochemical parameters related to cerebellum of rats.

Keywords: Cerebellum, Glutathione peroxidase, Motor activities, Oxidative stress markers

INTRODUCTION

In 21st century world, industrialisation of society and expansion of factories has vividly expanded the amount of pollutants and environmental pollutions. Environmental pollution due to arsenic is a major toxin and has become a major public health challenge in many countries such as Bangladesh, India, Nepal, Taiwan, Mongolia, Vietnam, Pakistan, China, Afghanistan, Argentina and USA [1]. Arsenic is mainly distributed in the environment through industries using wood preservatives, glass, semiconductors, mining waste, herbicides and pesticides [2]. Due to its increasing production and utilisation, arsenic is ubiquitously present in water, air and soil affecting occupational workers as well as general population [3].

Arsenic primarily enters into the body through direct consumption of drinking water from geological deposits by drilling of tube well [4]. World Health Organization (WHO) and other regulatory bodies have established a Maximum Contaminant Limit (MCL) of 10 µg/L inorganic arsenic (iAs) in drinking water for the safety of human health based on reducing cancer risk, but this limit is not considering endpoints other cancer end points such as digestive effects, reproductive effect [5]. Globally, more than 200 million people are affected by ground-water contaminated with arsenic concentration greater than 10 µg/L [6]. Arsenic affected area in West Bengal and Bihar region of the Ganga

Plain in India has extremely high arsenic contaminated water where arsenic concentration in water has been reported upto 1000 µg/L (i.e., 100 PPM) against the safe limit of 10 µg/L recommended by WHO [7,8]. High dosages iAs can cause death but chronic lower levels of arsenic results in serious health problems such as cancers and skin lesions [9]. Numerous experimental and epidemiological data provide evidence that acute and chronic arsenic exposure has been linked with numerous chronic indices in all human and animal organ systems and contributes to wide spectrum of diseases, especially diseases involving central nervous system [10-12].

Cerebellum is an important component of brain which is linked to control of posture, coordination of movements, gait and skilled voluntary movements. Some studies suggest that cerebellum is also involved in cognition, language and executive functioning [13]. Arsenic penetrates the Blood Brain Barrier (BBB) and accumulates in different regions of the brain such as cortex, cerebellum and hippocampus and increases the production of reactive oxygen species in brain which causes an imbalance between the antioxidant defense system levels and the production of free radicals to prompt neurotoxicity [14,15].

Arsenic induced oxidative stress causes neuronal injuries leading to neuronal cell death and impairment of brain functions because of its

high energy demand, high oxygen consumption and abundance of polyunsaturated lipid contents and have shown various neurological side-effects such as vertigo, impaired coordination of movements, uncontrolled motor learning, unsteady gait, loss of skilled voluntary movements, impaired learning, memory and concentration [16-18]. Reports in the literature also documented that arsenic-treated rats reduces neuronal viability in primary cultures of rat cerebellar neurons and showed alterations in locomotor behaviour and learning task [19,20].

These literatures indicate that arsenic has an impact on the cerebellum. There are minimal studies with a possible impact of arsenic on neurobehavioural and biochemical changes related with cerebellum [21,22]. Considering the lacunae, the present study aimed to investigate the impact of arsenic in cerebellum associated neurotoxicity by evaluating motor coordination, balance, locomotor and exploratory activities of rats with standard neurobehavioural procedure and also evaluated the biomarkers of oxidative stress.

MATERIALS AND METHODS

This experimental study was conducted in the Central Animal House at Sri Devaraj Urs Academy of Higher Education and Research (SDUAHER) from November 2019 to February 2020 for a period of 90 days. Full details of work plan with experimental animals were approved by Institutional Animal Ethics Committee (IAEC) (IAEC/PHARMA/SDUMC/2018-19/12a from SDUAHER, Kolar). All experiments were made according to guidelines established by the Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA) [23]. All efforts were made to minimise animal suffering and to reduce the number of animals used.

Inclusion criteria: Healthy male Sprague-Dawley rats having 150 to 180 grams in weight were included in the study.

Exclusion criteria: Rats with any signs/symptoms of illness as well as female rats were excluded from this study.

After two weeks of acclimatisation, all 16 rats were randomly divided into two groups of eight rats each.

Group I: Control, received normal saline.

Group II: Arsenic (as sodium arsenite at the doses of 50 PPM once in a day for 90 days through oral gavage).

Study Procedure

The dose of arsenic was chosen according to a previously published study [24]. Because of paucity in behavioural parameters and duration of exposure (90 days), the dose of present study was chosen on previously published report because the experimental animal model start appearing number of clinical and biochemical manifestation after 8-10 weeks of exposure.

Chemical: Sodium arsenite (NaAsO_2), purchased from LOBA Chemicals, was used in this study. All other reagents used in this experiment were of analytical grade and obtained from commercial sources.

Experimental animals: Male Sprague-Dawley rats free of specific pathogens were purchased from Biogen Attibele, Bangalore, India and kept in the central animal house of Sri Devaraj Urs Medical College, Kolar Karnataka, India. Rats were 10 to 12 weeks of age with a body weight varying from 150 to 180 grams and procured in well-maintained temperature ranging from 20°C-26°C with a 12 hours dark/light cycles. They were housed in polypropylene home cages confirming two rats per cage. Throughout the experimentation, rats were given free access to rodent feed pellets and water Ad-libitum.

Neurobehavioural Studies

After 90 days of treatment, all rats underwent behavioural tests. The test was performed in a separate room maintaining 20°C-26°C with noise and dust free room. Rats were acclimatised to the test room for 30 minutes prior to the testing. The tests were performed between 09:00 AM to 03:00 PM.

1. Open Field Test (OFT): OFT was used to assess locomotion and exploratory behaviours [25].

Equipment: Briefly, OFT consisted of an open-top box (90×90 cm) with 45 cm high walls to prevent the rats from escaping. The floor of arena was painted black. White lines were drawn on arena floor and divided into 36 square blocks and each block was 15 cm² long. The central part consisted of four squares in the center of the apparatus and marked with a red line. Video camera and a 60 W white light bulb were fixed above the arena.

Procedure: Each rat was gently placed in the center of arena and allowed freely to explore the arena for 300 seconds. At the end of exploration, rats were transferred to their respective home cages. After each trails, arena was cleaned with 70% ethanol.

The following parameters were assessed [26]:

- Number of square line crossed
- Number of rearing
- Number of wall-rearing
- Time spent in center
- Freezing time
- Number of grooming

Note: Line crossed was only considered if the rats cross one of the lines with all four paws. Freezing time was only considered if the rats remain static for more than six seconds.

2. Beam Walking Test (BWT): Motor coordination and balance of rats were assessed by measuring the rat's ability to traverse narrow beams to reach an enclosed safety platform [27].

Equipment: The Beam consisted of 150 cm long strips of wood, stretched between a blind and home end. It was 5 cm and 2.5 cm square thick, placed 50 cm above the floor. Just below the beam, a soft cloth was placed to prevent rats from injury in case of fall from the beam during experiment. An aversive stimulus (60 W light) was positioned near the blind end of the beam [27].

Procedure: Rats were individually kept at the blind end and allowed to walk on the beam. The animals' performances were recorded with a video-camera. After each trails, test apparatus was cleaned with 70% alcohol to remove any smell and residue.

The following parameters were assessed:

- Time taken to traverse the beam
- Number of foot slips
- Number of near fall
- Actual fall

Note: All neurobehavioural tests were assessed three times and mean of these values was taken.

Sample collection: Following the behavioural tests, all rats were fasted over-night except water. Rats were deeply anaesthetised by injecting Ketamine (92 mg/kg Body-weight) and Xylocaine (10 mg/kg Body-weight) intraperitoneally and sacrificed by cervical decapitation. Blood was drawn from retro-orbital plexus and collected in heparinised vials, centrifuged to collect serum and stored in -80°C until use. Brain was removed, rinsed in cold saline and cerebellum was separated quickly. Half of the cerebellum was quickly frozen in liquid nitrogen and stored at -80°C and used for biochemical analysis. Remaining half was stored at -80°C and used for arsenic estimation.

Estimation of Arsenic Concentration in Serum and Cerebellum of Rat

Arsenic concentration in serum and cerebellum were assessed according to the procedure reported by Ballentine R et al., [28]. A 200 mg of cerebellum was added in di-acid mixture which contains HNO_3 and HClO_4 in the ratio of 10:4, respectively. The sample was digested over a sand bath fume-hood chamber until a white gelatinous residue was formed. It was cooled and brought

to 20 mL of volume by adding distilled water. Aliquots of digested and undigested samples such as cerebellum and serum were used to estimate arsenic using Atomic Absorption Spectrophotometer (Thermo-Scientific iCE 3000 series AA Spectrophotometer). The values are expressed in $\mu\text{g/g}$ for tissue and ng/mL in serum.

Preparation of tissue homogenate for biochemical parameters: Each cerebellar tissue was homogenised individually in phosphate buffer Solution (pH 7.4) by using Glass-Teflon homogeniser. The cerebellar tissue segments were cut and fabricated in a 1:10 W/V (i.e., 1 gram of tissue in 10 mL of PBS) and homogenised for 10-15 minutes at 4°C. The homogenate was centrifuged at 15000 xg for 30 minutes at 4°C to remove debris. Clear supernatant was taken and stored at -80°C [29]. The supernatant obtained from the tissues was used to estimate biochemical parameters such as Malondialdehyde, Nitric Oxide and Glutathione Peroxidase spectrophotometrically [Table/Fig-1] [30-32].

Parameters	Methods	Units
Malondialdehyde (MDA)	Thiobarbituric acid Method (TBARs) [30]	Serum: nmol/mL Tissue: nmol/mg of protein
Nitric Oxide (NO)	Modified Griess's method [31]	Serum: nmol/mL Tissue: nmol/mg protein
Glutathione Peroxidase (GPx)	Procedure of Rotruck et al. [32]	Serum: mmols of GSH oxidised/min/mL Tissue: mmols of GSH oxidised/min/mg protein

[Table/Fig-1]: Oxidative stress parameters [30-32].
GSH: Reduced glutathione

STATISTICAL ANALYSIS

Statistical Package for the Social Sciences (SPSS) IBM, version 22.0 was used for analysis. Mean \pm SD was used for normally distributed data and the significance of difference was tested by independent t-test. Non normally distributed data were represented as Median (25th-75th Percentiles). Mann-Whitney U test were used for non parametric variables to check the difference in median between the groups. p-value <0.05 was considered statistically significant.

RESULTS

General toxicity: There was alteration in eating, drinking and defecation. Following three to five days of treatment with arsenic, we noticed a reduction in food and water intake for 15-20 days, thereafter consumption of food and water was almost similar compared to controls. After 45 days till decapitation, they reduced food and water intake. A few changes were also noticed in arsenic treated rats like alteration in texture of stool, bloated abdomen, restlessness.

Body weight, brain weight and arsenic concentration: Arsenic treated rats showed significant reduction in the body weight gain after 90 days when compared to body-weight gain of control rats after 90 days. There was also significant decrease in brain weight in arsenic-exposed rats compared to controls (p-value <0.001). Group II also increased arsenic concentration in serum and cerebellum of rats [Table/Fig-2].

Oxidative Stress (OS) markers: To check the impact of NaAsO₂ on OS- parameters, the activity of GPx and concentration of MDA and NO in serum and cerebellum of rats were assessed [Table/Fig-3].

Parameters	Group I (n=8)	Group II (n=8)	p-value
Initial body weight (grams)	225.38 \pm 6.78	229.63 \pm 5.98	0.303
Body weight after 90 days (grams)	317.25 \pm 6.99	286.87 \pm 6.6	0.001
Brain weight (grams)	2.17 \pm 0.19	1.36 \pm 0.12	0.001
Arsenic concentration in serum (ng/mL)	1.14 \pm 0.43	5.5 \pm 1.6	0.001
Arsenic concentration in cerebellum ($\mu\text{g/g}$)	0.65 \pm 0.29	2.76 \pm 0.56	0.001

[Table/Fig-2]: Comparison of differences in body weight, brain weight and arsenic concentration in Group I and Group II.
Values are Mean \pm SD. Independent t-test was done to calculate p-value. p<0.05 considered statistical significant

Oxidative stress parameters	Group I (n=8)	Group II (n=8)	p-value
Serum			
Malondialdehyde (nmol/mL)	8.81 (8.36-9.48)	16.84 (13.84-18.87)	0.001
Nitric-Oxide (nmol/mL)	17.66 (15.33-21.29)	33.79 (30.05-37.17)	0.001
Glutathione-Peroxidase (mmoles of GSH oxidised/min/mL)	15.16 (12.77-16.59)	6.89 (5.24-8.5)	0.001
Cerebellum			
Malondialdehyde (nmol/mg of protein)	3.02 (2.35-3.61)	7.98 (7.14-8.92)	0.001
Nitric-Oxide (nmol/mg of protein)	13.93 (11.0-16.16)	24.67 (21.4-28.22)	0.001
Glutathione-Peroxidase (mmoles of GSH oxidised/min/mg protein)	7.63 (7.08-9.19)	2.66 (1.19-3.86)	0.001

[Table/Fig-3]: Effect of Sodium-Arsenite on oxidative stress parameters.
Data is expressed as Median (25th-75th percentile). Mann-Whitney U test was used to calculate p-value. p<0.05 is considered significant difference between the groups

The current results documented that rats treated with arsenic (Group II) significantly increases the concentration of MDA and NO and decreased the activity of GPx in serum and cerebellum of rats in comparison with control rats. The neurobehavioural activities were measured in OFT and BWT in experimental rats.

Effect of arsenic on OFT parameters: OFT was performed to assess the effect of oral administration of NaAsO₂ on locomotion and exploratory behaviour. Group II rats significantly reduced the frequency of square line crossed, rearing, wall rearing and no of grooming in comparison with group I. On the other hand, freezing time increased significantly in group II, but there was no difference in grooming duration between the groups [Table/Fig-4].

Parameters	Group I (n=8) Median (25 th -75 th percentile)	Group II (n=8) Median (25 th -75 th percentile)	p-value
Number of square line crossed	128.0 (121.67-141.75)	27.34 (24.5-28.25)	0.001
Number of rearing	13.67 (11.84-16.17)	3.0 (2.67-3.33)	0.001
Number of wall-rearing	25.17 (22.33-27.33)	10.67 (8.84-12.5)	0.001
Time spent in center (Seconds)	5.34 (3.5-13.75)	0.0 (0.0-0.75)	0.001
Freezing time (Seconds)	47.84 (39.5-62.0)	224.33 (210.67-247.92)	0.001
Number of grooming	10.5 (9.75-11.92)	4.0 (3.42-5.08)	0.001
Duration of grooming (Seconds)	26.34 (21.42-28.83)	24.33 (19.25-26.83)	0.312

[Table/Fig-4]: Effect of arsenic on Open Field Test (OFT) parameters.
Values are Median (25th-75th Percentile). Mann-Whitney U test was used to check significance difference. The p-value <0.05 considered statistically significant

Effect of arsenic on Beam Walking Test (BWT) parameters: BWT was done to assess the effect of oral administration of NaAsO₂ on motor co-ordination and balance of rats [Table/Fig-5a,b].

Parameters	Group I (n=8) Median (25 th -75 th percentile)	Group II (n=8) Median (25 th -75 th percentile)	p-value
Time spent to cross the beam (seconds)	19.7 (14.4-25.1)	48.8 (36.7-60.6)	0.001
Number of foot slip	1.17 (0.25-2.25)	6.17 (5.42-7.67)	0.001
Number of near-fall	0.0 (0.00-0.25)	2.5 (1.75-3.25)	0.001
Actual fall	0.0 (0.0-0.0)	0.17 (0.0-0.33)	0.025

[Table/Fig-5a]: Effect of Beam Walking Test (BWT) parameters in 5 cm thick beam.
Values are Median (25th-75th Percentile). Mann-Whitney U test was used to check significance difference. The p-value <0.05 considered statistically significant

The current study indicated that group II took longer time to traverse the beam and also experienced more number of foot slips and near fall in 2.5 cm and 5 cm thick beam compared to group I. This may suggest that NaAsO₂ alters the motor co-ordination and balance.

All rats (control and arsenic treated) took more time to traverse 2.5 cm square thick beam as compared to 5 cm thick beam. And

also number of foot slips and near falls was more in 2.5 cm thick beam as compared to 5 cm thick beam.

Parameters	Group I (n=8) Median (25 th -75 th percentile)	Group II (n=8) Median (25 th -75 th percentile)	p-value
Time spent to cross the beam (seconds)	9.0 (7.25-10.0)	39.67 (33.67-46.83)	0.001
Number of foot slip	0	2.5 (1.84-3.0)	0.001
Number of near-fall	0	1.33 (1.0-2.42)	0.001
Actual fall	0	0 (0.0-0.25)	0.144

[Table/Fig-5b]: Effect of Beam Walking Test (BWT) parameters in 2.5 cm thick beam. Data are depicted as Median (25th-75th Percentile). Mann-Whitney U test was done to calculate p-value. p<0.05 considered statistically significant.

DISCUSSION

There is an increasing concentration of arsenic in the environment due to rapid industrialisation and urbanisation, which contaminate the water, air and soil. The uses of arsenic contaminants are toxic to humans and animals even at low (<10 µg/L) concentration [33]. The current finding reported that the administration of NaAsO₂ causes an impact on neurobehavioural and neurochemical alterations in rat's cerebellum.

The present results revealed that arsenic treated rats cause a reduction in body weight gain which is similar to the study conducted by Sárközi K et al., and Rodríguez VM et al., [34,35]. The deficit in body weight gain could be due to reduced food consumption or damage to the gastric mucosa that causes gastrointestinal alterations and may alter the texture of stool [36]. The current study also distinguished that there was a significant reduction in brain weight which may indicate arsenic has crossed the BBB and altered the brain weight and changes in brain weight could indicate oxidative damage.

It is documented in animal experiments that arsenic can cross BBB and invade the brain parenchyma [20]. In present study, arsenic concentration in serum and cerebellum significantly increased in arsenic-treated rats which might be a contributing factor to cerebellar neurotoxicity. The present study is parallel with the results of Guan H et al., and Markowski VP et al., [37,38]. However, the change in serum and cerebellum arsenic concentration in the current and previous study could be due to different durations and routes of exposure and also to different groups of rodents.

OS is considered a deciding factor that contributes and triggers the induction of arsenic toxicity and has solid relationship with accumulation of arsenic in many organs such as heart, kidneys, lungs, liver, muscles, spleen and brain. Its accumulation has a persistent potential to damage organs and tissues [39]. To study the toxic signs caused by arsenic and its effect on oxidative stress, we assessed MDA, NO and GPx in serum and cerebellum.

The results of the present study indicated that NaAsO₂ causes significant neurotoxicity as confirmed by increase in OS markers such as MDA and NO as well as decrease in GPx activity in the serum and cerebellum of rats. In the current study, the concentration of MDA and NO level increased significantly in cerebellum which indirectly reflects the body cells by the severity of free radicals attacks. The present results also points out a positive relationship between MDA and NO levels and arsenic concentration in serum and cerebellum of rats. Simultaneously, the oxidative damage parameters i.e., GPx found to be significantly reduced due to antioxidant enzyme consumption and the antioxidant enzyme; GPx had the ability to scavenge oxygen free radicals. These results may show that arsenic can induce oxidative stress in rats' cerebellum. These findings are consistent with the earlier reports indicating an increase in MDA and NO level and decrease in GPx activity in the cerebellum [10,40,41]. Ingested arsenic accumulates in different targeted tissues and can interfere with many biochemical reactions and alter numerous physiological activities in various organs of body [42]. To substantiate

the motor coordination, balance, locomotion and exploratory behaviour in arsenic induced rats, we performed BWT and OFT.

Rats treated with NaAsO₂ showed marked impairment in locomotor activities as evidenced by the decreased frequency of square line crossed and increased freezing time as addressed in OFT which was similar to the study conducted by Kaushal P et al., and Saritha S et al., [43,44]. The reduction in locomotion might be due to a cerebellar disorder because the cerebellum is especially linked to voluntary nervous system and can result improper coordination between the nervous system and the muscular junction and might also be due to the oxidative stress detected in the of arsenic treated cerebellum which decreased motility in open field [20,34,45]. However, the present study data is contradictory to the study done by Bikashvili T et al., and stated that there was no difference in locomotion between the arsenic exposed and control rodents [46].

In addition, we also evaluated the exploratory activities and found that there was significant reduction in exploratory activities such as rearing and wall-rearing in arsenic-treated rats, which is parallel to the study conducted by Saritha S et al., [44]. The reduction in rearing in arsenic treated rats may indicate weakening of the limb muscles and leads to motor impairment and balance of the body.

Damage to the cerebellar structures promotes impairment in motor tasks such as coordination of movements, balance, and timing to traverse the beam. The present study showed that NaAsO₂ treated rats showed a significant impairment in fine motor co-ordination and balance by increasing time to cross the beam, number of foot-slips and near fall which may indicate arsenic induced cerebellar neurotoxicity. This is in consonance with the study done by Kim H et al., and stated that arsenic treated mice reduced the motor coordination compared to control [47]. This impairment in motor dysfunction may be linked with cerebellar markers of lipid and DNA oxidation [45].

Limitation(s)

Less number of animals was used in each group, microscopic study and other biochemical parameters has not been evaluated, rats were given sodium Arsenite orally through oral gavage, hence further research can be carried out in different experimental animals with other routes of administration.

CONCLUSION(S)

The oral administration of sodium arsenite (NaAsO₂) at the doses of 50 PPM once in a day for 90 days showed reduction in body weight gain. The arsenic treated rats had high concentration of arsenic in serum and cerebellar tissue which indicates accumulation of arsenic in cerebellum of rats and may reduce the brain weight and probably could have increased the production of free radicals and might cause oxidative stress. The present study also confirms that reduction in locomotion and exploratory behaviours and impairment in motor coordination and balance which indicate arsenic induced cerebellar neurotoxicity. Further, molecular studies may confirm the cerebellar neurotoxicity which may strengthen the present study.

Acknowledgement

Authors would like to thank Dr. Sarala N Department of Pharmacology and Incharge animal house facility, Sri Devaraj Urs Medical College, Kolar for providing animal house to carry out the study. Also would like to thank Mr. Ravi Shankar, Statistician, Department of Community medicine, for his timely help.

REFERENCES

- Brinkel J, Khan MH, Kraemer A. A systematic review of arsenic exposure and its social and mental health effects with special reference to Bangladesh. *Int J Environ Res Public Health*. 2009;6(5):1609-19.
- Verma CP, Bhatia R, Pandit V, Ashawat MS. Arsenic induced diseases in human beings, their diagnosis & treatment. *J Chem Pharm Res*. 2016;8(1):13-22.

- [3] Chouhan S, Flora SJ. Arsenic and fluoride: Two major ground water pollutants. *Indian J Exp Biol*. 2010;48(7):666-78.
- [4] Ali H, Khan E, Ilaqi I. Environmental chemistry and ecotoxicology of hazardous heavy metals: Environmental persistence, toxicity, and bioaccumulation. *Journal of Chemistry*. 2019;2019:6730305.
- [5] Bloom MS, Surdu S, Neamtii IA, Gurzau ES. Maternal arsenic exposure and birth outcomes: A comprehensive review of the epidemiologic literature focused on drinking water. *Int J Hyg Environ Health*. 2014;217(7):709-19.
- [6] Naujokas MF, Anderson B, Ahsan H, Aposhian HV, Graziano JH, Thompson C, et al. The broad scope of health effects from chronic arsenic exposure: Update on a worldwide public health problem. *Environ Health Perspect*. 2013;121(3):295-302.
- [7] Nordstrom DK. Public health. Worldwide occurrences of arsenic in ground water. *Science*. 2002;296(5576):2143-45.
- [8] McCarty KM, Hanh HT, Kim KW. Arsenic geochemistry and human health in South East Asia. *Rev Environ Health*. 2011;26(1):71-78.
- [9] Kumar S, Dubey RS, Tripathi RD, Chakrabarty D, Trivedi PK. Omics and biotechnology of arsenic stress and detoxification in plants: Current updates and prospective. *Environ Int*. 2015;74:221-30.
- [10] Samad N, Jabeen S, Imran I, Zulfiqar I, Bilal K. Protective effect of gallic acid against arsenic-induced anxiety-/depression- like behaviours and memory impairment in male rats. *Metab Brain Dis*. 2019;34(4):1091-102.
- [11] Kuo CC, Moon KA, Wang SL, Silbergeld E, Navas-Acien A. The association of arsenic metabolism with cancer, cardiovascular disease, and diabetes: A systematic review of the epidemiological evidence. *Environ Health Perspect*. 2017;125(8):08700.
- [12] Cholanians AB, Phan AV, Ditzel EJ, Camerisch TD, Lau SS, Monks TJ. From the cover: Arsenic induces accumulation of α -Synuclein: Implications for synucleinopathies and neurodegeneration. *Toxicol Sci*. 2016;153(2):271-81.
- [13] Manto M. Toxic agents causing cerebellar ataxias. *Handb Clin Neurol*. 2012;103:201-13.
- [14] Punshon T, Davis MA, Marsit CJ, Theiler SK, Baker ER, Jackson BP, et al. Placental arsenic concentrations in relation to both maternal and infant biomarkers of exposure in a US cohort. *J Expo Sci Environ Epidemiol*. 2015;25(6):599-603.
- [15] Jomova K, Jenisova Z, Feszterova M, Baros S, Liska J, Hudecova D, et al. Arsenic: toxicity, oxidative stress and human disease. *J Appl Toxicol*. 2011;31(2):95-107.
- [16] Herrera A, Pineda J, Antonio MT. Toxic effects of perinatal arsenic exposure on the brain of developing rats and the beneficial role of natural antioxidants. *Environ Toxicol Pharmacol*. 2013;36(1):73-79.
- [17] Tolins M, Ruchirawat M, Landrigan P. The developmental neurotoxicity of arsenic: Cognitive and behavioural consequences of early life exposure. *Ann Glob Health*. 2014;80(4):303-14.
- [18] Piao F, Li S, Li Q, Ye J, Liu S. Abnormal expression of 8-nitroguanine in the brain of mice exposed to arsenic subchronically. *Ind Health*. 2011;49(2):151-57.
- [19] Namgung U, Xia Z. Arsenic induces apoptosis in rat cerebellar neurons via activation of JNK3 and p38 MAP kinases. *Toxicol Appl Pharmacol*. 2001;174(2):130-38.
- [20] Rodriguez VM, Carrizales L, Jimenez-Capdeville ME, Dufour L, Giordano M. The effects of sodium-Arsenite exposure on behavioural parameters in the rat. *Brain Research Bulletin*. 2001;55(2):301-18.
- [21] Adedara IA, Fabunmi AT, Ayenitaju FC, Atanda OE, Adebowale AA, Ajayi BO, et al. Neuroprotective mechanisms of selenium against arsenic-induced behavioural impairments in rats. *Neurotoxicology*. 2020;76:99-110.
- [22] Peruru R, Dodoala S. Therapeutic potential of diosmin, a citrus flavonoid against arsenic-induced neurotoxicity via suppression of NOX 4 and its subunits. *Indian J Pharmacol*. 2021;53(2):132-42.
- [23] Care V. CPCSEA guidelines for laboratory animal facility. *Indian J Pharmacol*. 2003;35:257-74.
- [24] Mishra D, Flora SJ. Differential oxidative stress and DNA damage in rat brain regions and blood following chronic arsenic exposure. *Toxicol Ind Health*. 2008;24(4):247-56.
- [25] Donatti AF, Soriano RN, Leite-Panissi CR, Branco LG, de Souza AS. Anxiolytic-like effect of hydrogen sulfide (H₂S) in rats exposed and re-exposed to the elevated plus-maze and open field tests. *Neurosci Lett*. 2017;642:77-85.
- [26] Fan SJ, Jiang H, Yang LJ, Liu X, Song J, Pan F. Effects of adrenergic agents on stress-induced brain microstructural and immunochemical changes in adult male Wistar rats. *Ann Anat*. 2011;193(5):418-24.
- [27] Silveira EMS, Kroth A, Santos MCQ, Silva TCB, Silveira D, Riffel APK, et al. Age-related changes and effects of regular low-intensity exercise on gait, balance, and oxidative biomarkers in the spinal cord of Wistar rats. *Braz J Med Biol Res*. 2019;52(7):e8429.
- [28] Ballentine R, Burford DD. Determination of metals. *Methods Enzymol*. 1957;3:1002-35.
- [29] Mohammadi MT, Amini R, Jahanbakhsh Z, Shekarforoush S. Effects of atorvastatin on the hypertension-induced oxidative stress in the rat brain. *Iran Biomed J*. 2013;17(3):152-57.
- [30] Esteribauer H, Cheeseman K. Determination of aldehydic lipid peroxidation products: Malonaldehyde on related aldehyde. *Free Radic Biol Med*. 1991;11:81-128.
- [31] Green LC, Wagner DA, Glogowski J, Skipper PL, Wishnok JS, Tannenbaum SR. Analysis of nitrate, nitrite, and [15N] nitrate in biological fluids. *Analytical Biochemistry*. 1982;126(1):131-38.
- [32] Rotruck JT, Pope AL, Ganther HE, Swanson AB, Hafeman DG, Hoekstra WG. Selenium: biochemical role as a component of glutathione peroxidase. *Science*. 1973;179(4073):588-90.
- [33] Jalaludeen AM, Ha WT, Lee R, Kim JH, Do JT, Park C, et al. Biochanin A Ameliorates Arsenic-induced hepato- and hematotoxicity in rats. *Molecules*. 2016;21(1):69.
- [34] Sárközi K, Horváth E, Vezér T, Papp A, Paulik E. Behavioural and general effects of subacute oral arsenic exposure in rats with and without fluoride. *Int J Environ Health Res*. 2015;25(4):418-31.
- [35] Rodríguez VM, Limón-Pacheco JH, Carrizales L, Mendoza-Trejo MS, Giordano M. Chronic exposure to low levels of inorganic arsenic causes alterations in locomotor activity and in the expression of dopaminergic and antioxidant systems in the albino rat. *Neurotoxicol Teratol*. 2010;32(6):640-47.
- [36] Klaassen CD. Heavy metals and heavy-metal antagonists. *The Pharmacological Basis of Therapeutics*. 1996;10:1851-75.
- [37] Guan H, Li S, Guo Y, Liu X, Yang Y, Guo J, et al. Subchronic exposure to arsenic represses the TH/TR β 1-CaMK IV signaling pathway in mouse cerebellum. *Int J Mol Sci*. 2016;17(2):01-16.
- [38] Markowski VP, Reeve EA, Onos K, Assadollahzadeh M, McKay N. Effects of prenatal exposure to sodium arsenite on motor and food-motivated behaviours from birth to adulthood in C57BL6/J mice. *Neurotoxicol Teratol*. 2012;34(2):221-31.
- [39] Sayed S, Ahsan N, Kato M, Ohgami N, Rashid A, Akhand AA. Protective effects of phyllanthus emblica leaf extract on sodium arsenite-mediated adverse effects in mice. *Nagoya J Med Sci*. 2015;77:145-53.
- [40] Goudarzi M, Amiri S, Nesari A, Hosseinzadeh A, Mansouri E, Mehrzadi S. The possible neuroprotective effect of ellagic acid on sodium arsenate-induced neurotoxicity in rats. *Life Sci*. 2018;198:38-45.
- [41] Durappanavar PN, Nadoor P, Waghe P, Pavithra BH, Jayaramu GM. Melatonin ameliorates neuropharmacological and neurobiochemical alterations induced by subchronic exposure to arsenic in wistar rats. *Biol Trace Elem Res*. 2019;190(1):124-39.
- [42] Irshad K, Rehman K, Akash MSH, Hussain I. Biochemical investigation of therapeutic potential of resveratrol against arsenic intoxication. *Dose Response*. 2021;19(4):15593258211060941.
- [43] Kaushal P, Kumar P, Dhar P. Ameliorative role of antioxidant supplementation on sodium-arsenite induced adverse effects on the developing rat cerebellum. *J Ayurveda Integr Med*. 2020;11(4):455-63.
- [44] Saritha S, Davuljigari CB, Kumar KP, Reddy GR. Effects of combined arsenic and lead exposure on the brain monoaminergic system and behavioural functions in rats: Reversal effect of MiADMSA. *Toxicol Ind Health*. 2019;35(2):89-108.
- [45] Polotow TG, Poppe SC, Vardaris CV, Ganini D, Guariroba M, Mattei R, et al. Redox status and neuro inflammation indexes in cerebellum and motor cortex of wistar rats supplemented with natural sources of Omega-3 fatty acids and Astaxanthin: Fish oil, krill oil, and algal biomass. *Mar Drugs*. 2015;13(10):6117-37.
- [46] Bikashvili T, Lordkipanidze T, Gogichashvili N, Pochkhidze N. Effect of arsenic exposure on behaviour of rats of various age groups. *Georgian Med News*. 2017;264:119-26.
- [47] Kim H, Lee D, Kim K. Combined exposure to metals in drinking water alters the dopamine system in mouse striatum. *Int J Environ Res Public Health*. 2021;18(12):01-11.

PARTICULARS OF CONTRIBUTORS:

1. PhD Scholar, Department of Anatomy, Sri Devaraj Urs Medical College, Kolar, Karnataka, India.
2. Professor and Head, Department of Anatomy, Sri Devaraj Urs Medical College, Kolar, Karnataka, India.
3. Professor and Head, Department of Biochemistry, Sri Devaraj Urs Medical College, Kolar, Karnataka, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Sridevi Nangali Srinivasa,
Professor and Head, Department of Anatomy, Sri Devaraj Urs Medical College,
Kolar, Karnataka, India.
E-mail: sridevins26@gmail.com

AUTHOR DECLARATION:

- Financial or Other Competing Interests: None
- Was Ethics Committee Approval obtained for this study? Yes
- Was informed consent obtained from the subjects involved in the study? NA
- For any images presented appropriate consent has been obtained from the subjects. NA

PLAGIARISM CHECKING METHODS: [\[Jain H et al.\]](#)

- Plagiarism X-checker: Oct 31, 2021
- Manual Googling: Jan 19, 2022
- iThenticate Software: Feb 18, 2022 (7%)

ETYMOLOGY: Author Origin

Date of Submission: **Oct 28, 2021**
Date of Peer Review: **Nov 30, 2021**
Date of Acceptance: **Jan 25, 2022**
Date of Publishing: **Mar 01, 2022**